Coenzyme Q\(_{10}\) therapy before cardiac surgery improves mitochondrial function and in vitro contractility of myocardial tissue

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Objectives: Previous clinical trials suggest that coenzyme Q\(_{10}\) might afford myocardial protection during cardiac surgery. We sought to measure the effect of coenzyme Q\(_{10}\) therapy on coenzyme Q\(_{10}\) levels in serum, atrial trabeculae, and mitochondria; to assess the effect of coenzyme Q\(_{10}\) on mitochondrial function; to test the effect of coenzyme Q\(_{10}\) in protecting cardiac myocardium against a standard hypoxia-reoxygenation stress in vitro; and to determine whether coenzyme Q\(_{10}\) therapy improves recovery of the heart after cardiac surgery.

Methods: Patients undergoing elective cardiac surgery were randomized to receive oral coenzyme Q\(_{10}\) (300 mg/d) or placebo for 2 weeks preoperatively. Pectinate trabeculae from right atrial appendages were excised, and mitochondria were isolated and studied. Trabeculae were subjected to 30 minutes of hypoxia, and contractile recovery was measured. Postoperative cardiac function and troponin I release were assessed.

Results: Patients receiving coenzyme Q\(_{10}\) (n = 62) had increased coenzyme Q\(_{10}\) levels in serum (P = .001), atrial trabeculae (P = .0001), and isolated mitochondria (P = .0002) compared with levels seen in patients receiving placebo (n = 59). Mitochondrial respiration (adenosine diphosphate/oxygen ratio) was more efficient (P = .012), and mitochondrial malondialdehyde content was lower (P = .002) with coenzyme Q\(_{10}\) than with placebo. After 30 minutes of hypoxia in vitro, pectinate trabeculae isolated from patients receiving coenzyme Q\(_{10}\) exhibited a greater recovery of developed force compared with those in patients receiving placebo (46.3% ± 4.3% vs 64.0% ± 2.9%, P = .001). There was no between-treatment difference in preoperative or postoperative hemodynamics or in release of troponin I.

Conclusions: Preoperative oral coenzyme Q\(_{10}\) therapy in patients undergoing cardiac surgery increases myocardial and cardiac mitochondrial coenzyme Q\(_{10}\) levels, improves mitochondrial efficiency, and increases myocardial tolerance to in vitro hypoxia-reoxygenation stress.
In the current era, patients referred for cardiac surgery have increasing numbers of risk factors, including previous angioplasty, myocardial failure, and old age. Age-associated cardiovascular changes adversely affect the preoperative state of the myocardium, rendering it more vulnerable to stresses as such as those incurred during cardiac surgery and resulting in higher operative mortality and an increased incidence of postoperative complications.2,3

Coenzyme Q10 (CoQ10) is a lipid-soluble antioxidant and a key component of the mitochondrial electron transport chain for adenosine triphosphate (ATP) production.4,5 Myocardial CoQ10 content is reduced by cardiac failure and aging.6-8 Further reductions can be caused by 3-hydroxy-3-methyl-glutaryl coenzyme A (HMG-CoA) reductase inhibitors, which have been shown to lower plasma and myocardial levels of CoQ10.9-12 Our previous experimental work has shown that CoQ10 has the ability to protect the myocardium against ischemia-reperfusion injury8 and against aerobic pacing stress, especially in elderly hearts.13 Previous clinical trials have suggested that treatment with CoQ10 before cardiac surgery might improve postoperative cardiac function and reduce myocardial structural damage.14-19 However, many of these trials were limited by insufficient CoQ10 dosage or low patient numbers or were not randomized, double-blinded, and placebo controlled. Although some of these studies demonstrated increased serum levels, incorporation of CoQ10 into the myocardium and mitochondria was not studied. Thus the aim of the present study was to perform a prospective, randomized, double-blind trial of preoperative high-dose CoQ10 therapy (300 mg/d) in patients undergoing elective cardiac surgery on bypass to measure the effect of high-dose CoQ10 therapy on the following: (1) CoQ10 levels in serum, atrial myocardium, and mitochondria (dose efficacy); (2) mitochondrial respiration; and (3) capacity to protect atrial trabeculae against a standard hypoxia-reoxygenation stress in vitro (primary end point). We also sought to assess the effect of CoQ10 therapy on postsurgical recovery of cardiac pump function, troponin release, and quality of life (secondary end points).

Methods

Patient Enrollment

Patients presenting for elective first-time coronary artery bypass graft or valve operations with cardiopulmonary bypass between May 1998 and July 2000 at the Alfred Hospital were eligible for inclusion in the study. Exclusion criteria were reoperation, urgent or emergency procedures, current therapy with warfarin or anti-oxidants, and recent myocardial infarction (≤6 weeks before the operation).

Patients undergoing elective coronary bypass surgery who provided informed consent were randomized (random numbers in blocks of 4) to receive either oral CoQ10 (300 mg/d) or placebo in a double-blinded fashion before the operation, commencing at the time they were placed on the preoperative waiting list. The study protocol was approved by The Alfred Hospital Human Ethics Committee.

Atrial Appendage Harvest and Dissection

Atrial appendages discarded during atrial cannulation before cardiopulmonary bypass were stored in bicarbonate buffer (NaCl, 125.8 mmol/L; KCl, 3.6 mmol/L; MgSO4, 0.6 mmol/L; NaH2PO4, 1.3 mmol/L; NaHCO3, 25.0 mmol/L; glucose, 11.2 mmol/L; and CaCl2, 2.5 mmol/L equilibrated with 95% O2 and 5% CO2 to pH 7.4) containing 30 mmol/L 2,3 butanediol monoxide. All trabeculae were dissected out for isolation of mitochondria, and up to 6 pectinate trabeculae (~1 mm in diameter, <7 mm in length) per patient were selected for isometric contraction experiments.

Mitochondrial Isolation and Respiration

Mitochondria were isolated from atrial trabeculae according to previously reported methods.20 Mitochondrial O2 consumption was measured by using a miniature, custom-designed, Clarke-type O2 electrode in a glass chamber maintained at 37°C. Digital data acquisition and consumption rate analyses were performed with customized software. Mitochondria were suspended in sucrose buffer (0.3 mol/L sucrose, 5 mmol/L KH2PO4, 5 mmol/L (3-(N-morpholino)propanesulfonic acid (MOPS), 1 mmol/L ethylenediamine tetraacetate acid, and 0.1% bovine serum albumin–fatty acid free) and were added to the glass chamber at a protein concentration of 1 mg/mL. Pyruvate-malate (2.5 mmol/L and 0.5 mmol/L, respectively) was used as a source of electrons for the respiratory chain, whereas 0.34 mmol/mL adenosine diphosphate (ADP) was used to release the accumulation of electrons and drive ATP synthesis at Complex V. The proton uncoupler carbonyl cyanide 3-trifluoromethoxyphenylhydrazone (FCCP) (1 μmol/L) was used to determine the maximum O2 consumption rate. Sodium cyanide (10 mmol/L) was used to block mitochondria-specific O2 consumption at the conclusion of each experiment.

CoQ10 and Malondialdehyde Assays

Total CoQ10 was measured by means of UV spectrophotometric high-performance liquid chromatography analysis after solvent extraction of serum or isolated membranes with a mixture of hexanes and ethanol (1:5:2). CoQ10 (5 μg per sample) was used as the internal standard to determine the extraction efficiency. Samples and standards were measured on Waters high-performance liquid chromatography at 280 nm (20 minutes’ run time) by using a 39:35:26 mixture of methanol, isopropanol, and acetoniitrile as the isocratic mobile phase through a C18 Radial Pak, 8 mm inner diameter, 4-μm particle column (Waters). CoQ6 and CoQ10 standards (Sigma) ranged from 0.25 to 4 μg and 0.1 to 1.0 μg, respectively. The areas under the peaks of the samples were compared with those of the standards to determine CoQ10 content. Values were adjusted to the efficiency of CoQ6 extraction and were expressed as micrograms of CoQ10 per milligrams of mitochondrial protein or micrograms of CoQ10 per milliliters of serum.

Malondialdehyde (MDA) formation in isolated mitochondrial membranes was measured spectrophotometrically as an index of the degree of lipid peroxidation. A commercial colorimetric assay (catalog no. 437634, Calbiochem) was used to specifically mea-
sure the condensation between one molecule of MDA with 2 molecules of N-methyl-2-phenylindole to yield a stable chromophore at 586 nm.

**Atrial Trabecular Function in Vitro**

Trabeculae were attached to organ bath force transducers and were stimulated electrically at 2 Hz in oxygenated bicarbonate buffer, as previously described. After baseline measurements of contractility, hypoxia was induced by replacing the 95% O2/5% CO2 in the organ bath with 95% N2/5% CO2. After 30 minutes of hypoxia (Po2, 45 ± 3 mm Hg), the trabeculae were reoxygenated, and contractile recovery was continuously measured. Posthypoxic recovery of developed force (at 30 minutes) was expressed as a percentage of the hypoxic value. Measurements were also made of resting force, time to peak contraction, and time to 50% relaxation of peak force.

**Clinical Procedures**

Blood samples were taken for the measurement of CoQ10 levels before and after therapy (in the operating room before cardiopulmonary bypass). Coronary bypass surgery was performed by using a standard on-pump surgical technique. Myocardial preservation was attained by means of either intermittent tepid blood cardioplegia with St Thomas’ potassium-magnesium solution containing 15 mmol/L aspartate at 20°C to 25°C (3 surgeons) or by means of cold crystalloid St Thomas’ cardioplegia at 4°C with topical cooling (3 surgeons). Blood samples were taken for measurement of troponin I levels 4 and 24 hours after admission to the intensive care unit.

**Hemodynamic Monitoring**

A pulmonary artery thermodilution catheter (Edwards Lifescience) was inserted before the operation for measurement of pulmonary capillary wedge pressure and cardiac output. Measurements were taken in the operating room before cardiopulmonary bypass and after bypass just before closure of the sternotomy and again in the intensive care unit 4 hours after cardiac surgery. Heart rate and mean arterial blood pressure were also recorded and used to calculate left ventricular stroke work index. Inotropic drug therapy was instituted according to standardized criteria.

**Quality-of-life Measures**

The Medical Outcome Study Short Form 36-item health status questionnaire (SF36) was used to assess the quality of life after the operation. The SF36 is a self-administered questionnaire that assesses 8 variables related to physical and mental state, including functional status, emotional and social well-being, and overall evaluation of health. The survey was sent to all patients, and replies were obtained in the first cohort of 60 patients at a median of 27 months. The physical function parameters were combined to produce a Physical Component Summary, and the mental function parameters were combined to produce a Mental Component Summary.

**Statistics**

Sample size calculations showed that to detect a 10% between-group difference in the posthypoxic recovery of pectinate trabeculae developed force with a power of 80% and a significance level of .05 would require 94 patients per group. To detect a 15% difference would require 48 patients per group. Variables are expressed as means ± SEM. Log transforms were used to normalize length of stay and troponin levels. The t test was used for between-group differences in patient demographics, operative variables, CoQ10 levels, trabecular contractile function, and quality-of-life scores. The χ2 and Fisher exact tests were used for categoric variables. Multivariable analysis was used to test for the influence of preoperative variables on recovery of trabecular contractile function and troponin I release and for the influence of preoperative and operative variables on length of hospital stay. Values are reported as means ± SEM unless otherwise stated. Statistical significance was defined as P < .05.

**Results**

**Clinical Variables**

One hundred twenty-one patients were enrolled in the study. There were no significant differences at baseline between the 2 treatment groups (Table 1). The mean length of treatment in each group was approximately 2 weeks. The operative procedures were similar in both groups (Table 2), being predominantly coronary artery bypass graft surgery or valve replacement. The 6 operating surgeons were distributed equally between the 2 groups (P = .85). There were no deaths in the hospital.

**In Vitro Results**

**CoQ10 levels.** At baseline, serum CoQ10 levels were similar in both groups (Table 3). After therapy, serum CoQ10 levels in the CoQ10 group were approximately 4 times greater than those in the placebo group (P = .001). In atrial myocardium the CoQ10 concentration was 2.5 times higher in the CoQ10 group than in the placebo group (P = .0001). The mitochondrial CoQ10 concentration was 2.4 times greater than those in the placebo group (>60%; 2, mildly impaired (ejection fraction 46% to 60%); 3, moderately impaired (ejection fraction 30% to 45%); 4, severely impaired (ejection fraction <30%). Cosp10, Coenzyme Q10; remote MI, myocardial infarction more than 2 months before surgical intervention; LV, left ventricular; ACE, angiotensin-converting enzyme.

*Median (50% confidence limits).*

<table>
<thead>
<tr>
<th>TABLE 1. Patient demographics and length of therapy</th>
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<tr>
<td></td>
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<tr>
<td>Age (y)*</td>
</tr>
<tr>
<td>Male (%)</td>
</tr>
<tr>
<td>Remote MI (%)</td>
</tr>
<tr>
<td>LV function grade</td>
</tr>
<tr>
<td>Hypertension (%)</td>
</tr>
<tr>
<td>Diuretic (%)</td>
</tr>
<tr>
<td>ACE inhibitor (%)</td>
</tr>
<tr>
<td>β-blocker (%)</td>
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<tr>
<td>Days of therapy</td>
</tr>
</tbody>
</table>
times greater in the CoQ10 group than in the placebo group ($P = .0002$).

**Mitochondrial respiration and lipid peroxidation.** Coupled oxygen consumption (state III) in isolated myocardial mitochondria was significantly lower in the CoQ10 group (49.0 ± 2.7 vs 72.2 ± 3.7 ng O atoms · min⁻¹ · mg⁻¹ protein, $P = .011$) than in the placebo group (Figure 1A). The amount of oxygen consumed after addition of 0.34 mmol/L ADP was used to calculate the ADP/O ratio after correction for contaminating adenosine monophosphate. This is a stoichiometric index of the amount of ATP produced (ADP consumed) per mole of oxygen used and is indicative of efficiency in ATP production. For the CoQ10 group, the ADP/O ratio (2.04 ± 0.2) was significantly higher than for placebo group mitochondria (1.17 ± 0.2; $P = .012$; Fig 1, B). Fig 1, C, demonstrates significantly lower accumulation of the lipid peroxidation product MDA in mitochondrial membranes in the CoQ10 group (0.9 ± 0.04 nmol/mg protein) than in the placebo group (1.6 ± 0.12 nmol/mg protein, $P = .002$).

**Trabecular contractile function.** At baseline, there were no differences in developed force, resting force, or time to 50% relaxation between the groups (Table 4). After hypoxia and reoxygenation, the recovery of developed force in the CoQ10 group (64.0% ± 2.9%) was greater than in the placebo group (46.3% ± 4.3%; $P = .001$, Figure 2). There were no between-group differences in resting force or time to 50% relaxation (Table 4). Multivariate analysis identified only treatment group as a predictor of recovery of developed force.

**Clinical Results**

**Hemodynamics.** Preoperative hemodynamic performance was similar for both groups, as were hemodynamic measurements in the operating room after discontinuation of bypass and stabilization (Table 5). Comparing postbypass values with prebypass values across both groups showed an increase in heart rate (median, 65 beats/min [50% confidence limit, 55-76 beats/min] to 90 beats/min [85-95 beats/min], $P = .001$), an increase in cardiac index (from 2.6 L · min⁻¹ · m⁻² [2.1-3.0 L · min⁻¹ · m⁻²] to 3.0 L · min⁻¹ · m⁻² [2.5-3.5 L · min⁻¹ · m⁻²]), and an increase in mean arterial pressure (110 ± 20 mm Hg to 130 ± 20 mm Hg, $P = .001$). The postoperative period was characterized by a rapid decrease in heart rate and mean arterial pressure, which stabilized at approximately 70 beats/min and 110 mm Hg, respectively. There were no between-group differences in the duration of mechanical ventilation or ICU stay.

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**TABLE 2. Operative variables**

<table>
<thead>
<tr>
<th>CABG alone (%)</th>
<th>Placebo (n = 59)</th>
<th>CoQ10 (n = 62)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>93</td>
<td>87</td>
<td>.43</td>
<td></td>
</tr>
</tbody>
</table>

| Mean grafts   | 2.9 ± 0.1      | 3.2 ± 0.1      | .17     |
| Meantime (min)* | 91 (70-107)   | 91 (75-106)   | .61     |
| CC time (min)* | 57 (42-73)    | 56 (47-75)    | .77     |
| TBC           | 39 (66%)       | 43 (69%)       | .85     |

**TABLE 3. Serum, atrial myocardium, and mitochondrial CoQ10 levels**

<table>
<thead>
<tr>
<th>CoQ10 measure</th>
<th>Placebo</th>
<th>CoQ10</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum baseline*</td>
<td>0.38 ± 0.02</td>
<td>0.39 ± 0.02</td>
<td>.98</td>
</tr>
<tr>
<td>Serum after therapy</td>
<td>0.42 ± 0.03</td>
<td>1.59 ± 0.06</td>
<td>.001</td>
</tr>
<tr>
<td>Myocardium after therapy†</td>
<td>17.2 ± 1.4</td>
<td>43.7 ± 3.1</td>
<td>.0001</td>
</tr>
<tr>
<td>Mitochondria after therapy‡</td>
<td>4.04 ± 0.71</td>
<td>9.53 ± 0.90</td>
<td>.0002</td>
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</tbody>
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*CoQ10, Coenzyme Q 10; CABG, coronary artery bypass grafting; CPB, cardiopulmonary bypass; CC, crossclamp; TBC, tepid blood cardioplegia.

†Placebo, n = 56; CoQ10, n = 51.

‡Placebo, n = 10; CoQ10, n = 10.

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*The normal range of serum CoQ10 in healthy individuals is 0.5 to 1.0 μg/mL. CoQ10 units are given for serum as micrograms per milliliter, for atrial myocardium as micrograms per gram of wet weight, and for atrial mitochondria as micrograms per milligram of protein.

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**Figure 1. A, Coupled O₂ consumption state III respiration in isolated human mitochondria during oxidation of pyruvate (2.5 mmol/L) plus malate (0.5 mmol/L) at 37°C. B, The efficiency of energy production expressed as a stochiometric ratio of ADP consumed during ATP production per atom of oxygen consumed. C, The concentration of the lipid peroxidation product MDA in isolated human mitochondria.**
TABLE 4. In vitro contractile function of isolated atrial trabeculae

<table>
<thead>
<tr>
<th></th>
<th>Placebo (n = 20)</th>
<th>CoQ10 (n = 26)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Developed force</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline (g)</td>
<td>1.02 ± 0.12</td>
<td>1.16 ± 0.07</td>
<td>.31</td>
</tr>
<tr>
<td>After hypoxic recovery</td>
<td>46.3 ± 4.3</td>
<td>64.0 ± 2.9</td>
<td>.001</td>
</tr>
<tr>
<td>Resting force</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline (g)</td>
<td>0.4 ± 0.04</td>
<td>0.5 ± 0.06</td>
<td>.18</td>
</tr>
<tr>
<td>After hypoxic recovery</td>
<td>92.0 ± 6.2</td>
<td>86.7 ± 2.5</td>
<td>.49</td>
</tr>
<tr>
<td>Time to 50% relaxation</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline (ms)</td>
<td>104.2 ± 5.9</td>
<td>104.5 ± 3.7</td>
<td>.96</td>
</tr>
<tr>
<td>After hypoxic recovery</td>
<td>89.1 ± 7.1</td>
<td>85.2 ± 2.4</td>
<td>.85</td>
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</table>

CoQ10: Coenzyme Q10.

m⁻² [2.7-3.4 L · min⁻¹ · m⁻²], P = .001); a reduction in stroke work index (from 31 g.m/m² per beat [29-39 g.m/m² per beat] to 29 g.m/m² per beat [23-34 g.m/m² per beat], P = .003), and a reduction in systemic vascular resistance (from 1187 dynes/s [993-1428 dynes/s] to 886 dynes/s [773-1012 dynes/s], P = .001). However, there were no between-group differences in postbypass versus prebypass values, except for a decrease in systemic vascular resistance that was greater in the placebo group than in the CoQ10 group (P = .029). We tested for an effect of cardioplegia type on postbypass to prebypass hemodynamics. There was no effect on any of the parameters measured. Also, 4 hours after surgical intervention, there was no difference between treatment groups in any of the individual hemodynamic measurements. The incidence of inotropic drug use in the 2 groups was not significantly different (CoQ10 group, 24%; placebo group, 33%; P = .39).

Troponin I release. Troponin I release measured 4 hours after the operation was 43.3 ± 1.2 µg/L in the CoQ10 group and 44.2 ± 1.2 µg/L in the placebo group (P = .87), and after 24 hours, it was 14.1 ± 1.2 µg/L in the CoQ10 group and 13.6 ± 1.2 µg/L in the placebo group (P = .64). Univariate analysis of the determinants of troponin release over the 2 time points indicated significant influences of surgeon’s technique and operation type but not age or CoQ10 treatment (Table 6). It was not possible to separate the effect of surgeon’s technique from cardioplegia type because each surgeon used only one type of cardioplegia: surgeons 1, 4, and 5 (n = 48) used crystalloid cardioplegia, and surgeons 2, 3, and 6 (n = 73) used blood cardioplegia. Troponin release was significantly higher (P < .0001) in the cold crystalloid cardioplegia user group (35.0 ± 1.2 µg/L) than in the tepid blood cardioplegia user group (17.2 ± 1.2 µg/L). The 2 types of cardioplegia were equally distributed between the 2 treatment groups and were therefore unlikely to have influenced the CoQ10 effect. Multivariate analysis showed significant influences on troponin I release of surgeon’s technique, operation type (coronary bypass versus valve procedure), and bypass time.

Length of hospital stay. The median postoperative length of stay in the CoQ10 group was 7.0 days (interquartile range, 5.0-9.0 days), and in the placebo group, the median stay was 6.0 days (interquartile range, 5.0-8.0 days). Significant univariate predictors of postoperative hospital length of stay were age (r² = 0.07, P = .0035), cardiopulmonary bypass time (r² = 0.11, P = .0002), and crossclamp time (r² = 0.08, P = .0013) but not CoQ10 treatment (r² = 0.002, P = .58). Multivariate analysis showed only age (P = .0027) as a predictor of length of hospital stay.

Late Follow-up
At a median follow-up time of 37 months (interquartile range, 25-39 months), there were 5 late deaths: 4 (3 non-cardiac and 1 cardiac) in the CoQ10 group and 1 (non-cardiac) in the placebo group (P = .16). Quality-of-life assessment (SF36 questionnaire) of the first 60 patients at a median of 27 months (93% complete) showed a significantly better (+13%) Physical Component Summary in the CoQ10 group (CoQ10 group, 47.5 ± 1.6; placebo group, 42.0 ± 2.1; P = .046 [higher SF36 scores indicate a greater quality of life]). No significant difference was evident in the Mental Component Summary (CoQ10 group, 51.2 ± 1.3; placebo group, 51.3 ± 1.1; P = .92).

Discussion
Cardiac surgery involves multiple stresses to the myocardium that contribute to cellular energy depletion. In the intraoperative period there is ischemia-reperfusion injury caused by aortic crossclamping. In the postoperative period there is hypoxia caused by respiratory insufficiency and aerobic stress (high oxygen-demand stress) caused by post-
CoQ10 was greatest in these elderly tissues.13 We have also shown that isolated human myocardial trabeculae obtained from patients aged 34 to 89 years are protected against ischemia-reperfusion injury when treated with CoQ10.14,24,25 Free radicals26 and calcium overload.27 We have previously reported that isolated human myocardial trabeculae exhibited an increased CoQ10 concentration in the CoQ10 group compared with the placebo group and had greater efficiency of oxygen consumption during energy production. The higher ratio of ADP:O seen in the CoQ10 group compared with the placebo group demonstrated that CoQ10 pretreatment afforded preservation of more efficient oxidative phosphorylation (ATP production), which is required to support in vitro posthypoxic contractile function and postoperative pump function. Associated with these effects, mitochondrial membranes from the CoQ10 group contained reduced levels of the lipid peroxidation product MDA compared with placebo, which indicates increased resistance to oxidative injury consistent with higher levels of CoQ10 in these membranes. These effects demonstrate the multiple molecular roles of CoQ10, in particular as an electron carrier, transferring electrons from nicotinamide adenine dinucleotide (NADH) dehydrogenase and succinate dehydrogenase to the cytochromes in the inner mitochondrial membrane during oxidative phosphorylation and as a free radical scavenger that inhibits lipid peroxidation.14,24

Myocardial Effects of CoQ10 Pretreatment

Mitochondria isolated from atrial trabeculae obtained from patients aged 34 to 89 years are also protected against ischemia-reperfusion injury when pre-exposed to CoQ10 suspension in vitro.8 Moreover, in this previous study myocardial tissue from elderly individuals showed increased susceptibility to injury, and the protective effect of CoQ10 was greatest in these elderly tissues. These in vitro effects of CoQ10 followed rapid uptake of CoQ10 into sarcolemmal and mitochondrial membranes. We therefore postulated that sustained oral administration of CoQ10 to cardiac patients would increase myocardial and mitochondrial CoQ10 content, improve energy production, and confer cellular protection against oxidative stress. These benefits at a myocardial level might be detectable in terms of improvements in cardiac function and rate of recovery in the postoperative period.

In the present study the two treatment groups were comparable in terms of preoperative risk factors, and both were elderly (Table 1). Both treatment groups were subjected to the same intraoperative stresses in terms of duration of aortic crossclamping time and cardiopulmonary bypass and extent of surgical procedure.

### TABLE 6. Determinants of troponin I release over the first 24 hours postoperatively

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Univariate predictors</th>
<th>Multivariate predictors</th>
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<td></td>
<td></td>
<td></td>
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<tr>
<td>Univariate predictors</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bypass time</td>
<td>.013</td>
<td></td>
</tr>
<tr>
<td>Surgeon’s technique</td>
<td>.017</td>
<td></td>
</tr>
<tr>
<td>Operation type (valve vs CABG only)</td>
<td>.041</td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td>.056</td>
<td></td>
</tr>
<tr>
<td>CoQ10 treatment</td>
<td>.83</td>
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<tr>
<td>Multivariate predictors</td>
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<td>Bypass time</td>
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<td>Surgeon’s technique</td>
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<tr>
<td>Operation type</td>
<td>.074</td>
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</tr>
</tbody>
</table>

CABG, Coronary artery bypass grafting; CoQ10, coenzyme Q10.
the collapse of mitochondrial proton-motive force and membrane potential, leading to the disruption of ionic homeostasis and oxidative phosphorylation in cell death signaling pathways, particularly after ischemia and reperfusion.\textsuperscript{29} Protection afforded by CoQ\textsubscript{10} from oxidative inactivation of creatine kinase and other key proteins during reperfusion is crucial in preserving energy metabolism and cardiac performance.\textsuperscript{14-19}

Atrial trabeculae isolated from CoQ\textsubscript{10}-pretreated patients subjected to hypoxia-reoxygenation in vitro demonstrated greater recovery of developed force compared with placebo. This greater capacity for recovery of contractile function was associated with increased CoQ\textsubscript{10} content in serum and myocardium after 2 weeks of oral CoQ\textsubscript{10} therapy. Cardiac troponin I release was strongly influenced by the different surgeon’s techniques, the type of operation, and the duration of cardiopulmonary bypass but not by preoperative CoQ\textsubscript{10} therapy.

Of interest was an indication of improved subjective assessment of physical quality of life (+13\%) in the CoQ\textsubscript{10} group compared with the placebo group when the first cohort of 60 patients was followed up 22 months postoperatively. However, these results should be interpreted with caution because of the fact that quality-of-life data were missing on 7\% of the patients and that subjective improvement in physical quality of life does not necessarily indicate improved cardiac pump function.

**Previous CoQ\textsubscript{10} Trials in Cardiac Surgery**

Previous randomized studies of CoQ\textsubscript{10} treatment in cardiac surgery have shown significant effects but are limited by small patient numbers (\textless 50).\textsuperscript{15-20} Of these, 3 studies\textsuperscript{15,17,18} showed an improvement in postoperative hemodynamics in CoQ\textsubscript{10}-treated patients. Zhou and colleagues\textsuperscript{14} showed that CoQ\textsubscript{10} treatment significantly reduced plasma levels of MDA and creatine kinase (cardiac isoenzyme). Chen and associates\textsuperscript{19} showed improved preservation of myocardial ultrastructure in CoQ\textsubscript{10}-treated patients. However, Taggart and coworkers\textsuperscript{16} reported no benefit of oral CoQ\textsubscript{10} pretreatment in a study of 20 randomized patients undergoing CABG who received 600 mg of CoQ\textsubscript{10} or placebo for 12 hours before surgical intervention. This latter result can be explained in terms of insufficient preoperative treatment time. Data from our preliminary studies indicated that despite a reasonably rapid increase in the blood CoQ\textsubscript{10} level within 24 hours, approximately 1 week of treatment was required to show a significant increase in human myocardial CoQ\textsubscript{10} concentration.

**Limitations of the Study**

There was no difference between groups in hemodynamic measurements made at rest in the operating room before or after cardiopulmonary bypass apart from a greater decrease in systemic vascular resistance (before vs after bypass) in the placebo group, which is probably a chance finding. Also, 4 hours after the operation, there were no between-group differences in hemodynamic measurements at rest. In the early postoperative period, any downward changes in cardiac function tend to be corrected by adjustments made in inotropic drug use by intensive care staff in response to perceived clinical need. Thus a useful indicator of major differences in postoperative function is inotropic drug use. However, this might be influenced by therapeutic aggressiveness by intensive care unit staff. We attempted to minimize therapeutic variability in our study by introducing fixed criteria for initiation of inotrope therapy in the intensive care unit. There was a small but nonsignificant difference in use of inotropic drugs in the 2 treatment groups (CoQ\textsubscript{10} group, 25%; placebo group, 33\%). Sample size calculations showed that it would require a sample size of 530 per group for such a difference to reach significance.

There was no effect of CoQ\textsubscript{10} on hemodynamics before or after surgical intervention. If transesophageal echocardiography had been available during the trial, pressure-volume loop measures would have been valuable. Reduction in postoperative release of troponin I would have been persuasive evidence of reduced intraoperative myocardial damage. However, any ability of CoQ\textsubscript{10} to significantly reduce the release of troponin I was overshadowed by the confounding influences of cardiopulmonary bypass duration, operation type, and between-surgeon variation in techniques of surgical intervention and myocardial preservation. Thus greater patient numbers would be required to ensure sufficient statistical power to specifically overcome these confounding factors.

**Conclusions and Clinical Implications**

The important finding of the present study is the ability of orally administered CoQ\textsubscript{10} to increase CoQ\textsubscript{10} levels in human myocardium and mitochondria. The beneficial action of augmented CoQ\textsubscript{10} levels involves increased protection of mitochondria and myofilaments against oxidative stress, with the consequent maintenance of efficient energy production and improved contractile recovery after hypoxia-reoxygenation stress in vitro. To directly test the effect of CoQ\textsubscript{10} on key postoperative patient outcomes, including more direct measures of recovery of cardiac performance and length of hospital stay, requires a well-resourced, large, multicenter trial with adequate statistical power to detect changes in clinically important end points.

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References


