Pretreatment with phenoxybenzamine attenuates the radial artery's vasoconstrictor response to α -adrenergic stimuli

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Background: Although the radial artery bypass conduit has excellent intermediate-term patency, it has a proclivity to vasospasm. We tested the hypothesis that brief pretreatment of a radial artery graft with the irreversible adrenergic antagonist phenoxybenzamine attenuates the vasoconstrictor response to the vasopressors phenylephrine and norepinephrine compared with the currently used papaverine/lidocaine.

Methods: Segments of human radial artery grafts were obtained after a 30-minute intraoperative pretreatment with a solution containing 20 mL of heparinized blood, 0.4 mL of papaverine (30 mg/mL), and 1.6 mL of lidocaine (1%). The segments were transported to the laboratory and placed into a bath containing Krebs-Henseleit solution and 10, 100, or 1000 $\mu\text{mol/L}$ phenoxybenzamine or vehicle. The segments were tested in organ chambers for contractile responses to increasing concentrations of phenylephrine and norepinephrine (0.5-15 $\mu\text{mol/L}$).

Results: Contractile responses to 15 $\mu\text{mol/L}$ phenylephrine in control radial artery segments averaged $44.2\% \pm 9.1\%$ of the maximal contractile response to 30 mmol/L KCl. Papaverine/lidocaine modestly attenuated contraction to 15 $\mu\text{mol/L}$ phenylephrine ($32.1\% \pm 5.9\%$; $P = .22$), but 1000 $\mu\text{mol/L}$ phenoxybenzamine completely abolished radial artery contraction ($-7.2\% \pm 4.4\%$; $P < .001$). The effect of 10 and 100 $\mu\text{mol/L}$ phenoxybenzamine on attenuating vasocontraction was intermediate between 1000 $\mu\text{mol/L}$ phenoxybenzamine and papaverine/lidocaine. Responses to 15 $\mu\text{mol/L}$ norepinephrine in control radial artery segments averaged $54.7\% \pm 7.5\%$ of maximal contraction to 30 mmol/L KCl. Papaverine/lidocaine modestly attenuated the contraction response of radial artery segments ($35.6\% \pm 5.1\%$; $P = .04$). In contrast, 1000 $\mu\text{mol/L}$ phenoxybenzamine showed the greatest attenuation of norepinephrine-induced contraction ($-10.5\% \pm 2.0\%$; $P < .001$).

Conclusions: A brief pretreatment of the human radial artery bypass conduit with 1000 $\mu\text{mol/L}$ phenoxybenzamine completely attenuates the vasoconstrictor responses to the widely used vasopressors norepinephrine and phenylephrine. Papaverine/lidocaine alone did not block vasoconstriction to these α -adrenergic agonists.

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Dissatisfied with the pathologic changes noted in saphenous vein bypass grafts in coronary artery bypass surgery, Carpentier and colleagues,¹ in 1973, advocated the use of the radial artery (RA) as a bypass conduit in the aorta-coronary position. They noted excellent handling qualities and length. Two years later, enthusiasm for the RA as a bypass conduit waned because of the reported high frequency of graft occlusion that was attributed to vasospasm.² By 1976, the use of RA grafts was completely abandoned. However, Acar and colleagues³ revisited the use of the RA after several patients in Carpentier and associates' original series were found to have patent, disease-free RA grafts after 15 years. In a subsequent series of patients, Acar and colleagues found 1- and 5-year patency rates to be 92% and 83%, respectively.⁴ Others have corroborated these excellent short-term and intermediate-term patency rates of the RA conduit.⁵⁻¹⁰ Important to the revival of the RA as a bypass conduit was the "no-touch" technique of harvesting a pedicled RA; avoiding endothelial damage by refraining from using intraluminal probes; using vasodilators such as papaverine, lidocaine, and nitroglycerin in soaking solutions; and administering parenteral calcium channel blockers after surgery. However, the use of the RA is not universally accepted, possibly because of reports of vasospasm and myocardial hypoperfusion in the immediate postoperative period despite the use of intraoperative papaverine and postoperative calcium channel blockers.^{11,12}

In 2000, Taggart and colleagues¹³ reported that phenoxybenzamine (PBZ), a noncompetitive and irreversible α -adrenergic receptor antagonist, blocked the contraction of the ex vivo human RA in response to epinephrine. Taggart and associates pretreated the RA with a soaking solution containing 100 mg of PBZ in 50 mL of heparinized blood (5.9 mmol/L PBZ) and tested the RA segments within 1 hour of treatment. Subsequently, Velez and colleagues¹⁴ demonstrated that in canine RA a much lower concentration of PBZ (1 μ mol/L) blocked the contractile response to increasing concentrations of norepinephrine (NE) and phenylephrine (PE) at 2, 24, and 48 hours after treatment. NE and PE are most often used for hemodynamic support in the postoperative period and are used during off-pump coronary bypass operations to treat hypotension caused by cardiac elevation and manipulation to visualize posterior and lateral target vessels. In addition, Velez and associates¹⁴ demonstrated that 1 μ mol/L papaverine, a vasodilator most often given intraluminally after the RA graft is harvested, did not attenuate the vasospasm associated with α -adrenergic stimuli. In this study, we determined the optimal concentration of PBZ in a buffered solution used to soak and pretreat RA segments harvested from patients undergoing coronary artery bypass graft surgery. The RA graft is typically har-

vested with the surrounding musculofascial pedicle, which may impede exposure of the vessel to agents designed to attenuate vasoconstriction, such as papaverine/lidocaine and PBZ. Therefore, we also tested the hypothesis that performing an incision through the musculofascial tissue (fasciotomy) would allow greater exposure of the graft to papaverine/lidocaine and PBZ and thereby increase the efficacy of these agents.

Materials and Methods

Surgical Preparation

RA segments were obtained from unused portions of RA conduits of patients having elective coronary artery bypass grafting with or without cardiopulmonary bypass at the Crawford Long Hospital of Emory University. A modified Allen's test was performed to assess the adequacy of preoperative collateral circulation to the hand.¹⁵ The RA was harvested with its pedicle containing the venae comitantes, perivascular fat, and areolar tissue (no fasciotomy) by using a no-touch technique. Branches of the RA were ligated with vascular clips. A subset of the RA grafts had the musculofascial tissue incised (with fasciotomy) to expose the areolar tissue adjacent to the graft. The RA was then placed in a solution containing 20 mL of heparinized blood, 1.6 mL of 1% lidocaine, and 0.4 mL of papaverine (30 mg/mL) for approximately 30 minutes at room temperature. The RA graft was flushed intraluminally with the blood/papaverine/lidocaine solution at the beginning and at 15 minutes of the soaking period to ensure exposure of the intimal PBZ. Before its placement in the aorta-coronary position, a small segment of the RA was obtained and immediately placed in Krebs-Henseleit (K-H) buffer (118 mmol/L NaCl, 4.7 mmol/L KCl, 1.2 mmol/L KH_2PO_4 , 1.2 mmol/L MgSO_4 , 2.5 mmol/L CaCl_2 , 12.5 mmol/L NaHCO_3 , and 10 mmol/L glucose) at 4°C, pH 7.4 and transported to our Cardiothoracic Research Laboratory.

Experimental Protocol

The RA segment with or without fasciotomy was placed into K-H buffer (pH 7.4) at 25°C with 10, 100, or 1000 μ mol/L PBZ or vehicle. The RA was flushed intraluminally twice with this solution, once at the beginning and once at the end of a 30-minute incubation period, which approximates the time between RA harvest and placement in the aorta-coronary position. In addition, control RA segments were obtained before intraoperative pretreatment of the conduit with the papaverine/lidocaine solution and received no other treatment. The segments were prepared for placement in organ bath chambers by carefully skeletonizing them in cold K-H buffer and cutting them into rings 3 to 5 mm in length. The rings were then mounted on stainless-steel hooks, connected to FT-03 force displacement transducers, and placed into Radnoti organ chambers (Radnoti Glass, Monrovia, Calif) containing 7 mL of oxygenated (95% oxygen/5% carbon dioxide) K-H buffer at 37°C and pH 7.4. Indomethacin (10 μ mol/L) was added to the buffer to block responses to endogenous prostanoids. The rings were stabilized for 1 hour with frequent buffer changes and set to a predetermined tension that allowed 75% of maximal contraction to 30 mmol/L KCl.

The rings were then incubated with increasing concentrations of PE (0.5-15 $\mu\text{mol/L}$) or NE (0.5-15 $\mu\text{mol/L}$). After the highest concentration of α -adrenergic agent was achieved, 30 mmol/L KCl was added to the bath to quantify the maximal non-receptor-mediated constriction. In randomly selected vessels, the integrity of the RA endothelium was also tested for its receptor-dependent relaxation response to incremental concentrations of acetylcholine (ACh), a stimulator of nitric oxide synthase. The rings were precontracted with the thromboxane A_2 mimetic U46619 (1.4 nmol/L) and then exposed to increasing concentrations of ACh (1 nmol/L to 11.7 $\mu\text{mol/L}$) in the presence of 10 $\mu\text{mol/L}$ indomethacin.

The changes in isometric force were quantified by using an analog-to-digital converter sampling at 2 Hz. The responses were analyzed with a Windows-based videographics program (SPECTRUM; Wake Forest University, Winston-Salem, NC). The force of contraction elicited by exposure to increasing concentrations of PE and NE was expressed as a percentage of the maximal contraction generated by KCl in each ring. The degree of relaxation after exposure to ACh was expressed as the percentage tension reduction from the maximal force of contraction obtained from U46619.

Chemicals

The following drugs were purchased from the Sigma Chemical Company (St Louis, Mo): acetylcholine chloride, KCl, NE, L-phenylephrine hydrochloride, K-H buffer, calcium chloride, and sodium bicarbonate. PBZ hydrochloride (Dibenzylamine) was a gift from SmithKline Beecham Pharmaceuticals (Collegeville, Pa).

Statistical Analysis

Data were analyzed for significance by using a 1-way analysis of variance comparing the control, papaverine/lidocaine, and PBZ groups at each concentration of NE and PE. If a significant difference between groups was assigned by analysis of variance, a post hoc Student-Newman-Keuls test was applied to locate the source of differences. All data are reported as mean $\pm$ SEM.

Results

PE caused a concentration-dependent vasoconstriction in control RA (n = 10) segments; the contraction achieved at the maximal concentration of PE (15 $\mu\text{mol/L}$) averaged $44.2\% \pm 9.1\%$ of the KCl response (Figure 1). Pretreatment of the RA in papaverine/lidocaine solution (n = 32) did not significantly attenuate the concentration-dependent contraction responses to PE. Contraction at the highest concentration of PE was reduced by only 27% of control vessels ($32.1\% \pm 5.9\%$; $P = .22$). In contrast, PBZ in addition to papaverine/lidocaine attenuated the vasoconstriction to PE in a dose-dependent manner (Figure 1). At the highest concentration of PE used (15 $\mu\text{mol/L}$), the vasoconstriction response was attenuated by 63% of control at 10 $\mu\text{mol/L}$ PBZ (n = 23; $16.5\% \pm 4.3\%$; $P = .02$), by 80% of control at 100 $\mu\text{mol/L}$ PBZ (n = 28; $8.7\% \pm 5.1\%$; $P = .003$), and by 116% of control at 1000 $\mu\text{mol/L}$ PBZ (n = 22; $-7.2\% \pm 4.4\%$; $P < .001$).

Figure 1. Radial artery vasoconstriction responses to increasing concentrations of phenylephrine, with or without pretreatment with 3 different concentrations of phenoxybenzamine (PBZ). PBZ attenuates vasoconstriction to phenylephrine in a concentration-dependent manner. PBZ, Phenoxybenzamine pretreatment; Pap/Lido, papaverine/lidocaine pretreatment; control, no pretreatment. * $P < .05$, 10^{-3} mol/L PBZ versus 10^{-4} and 10^{-5} mol/L PBZ, Pap/Lido, and control. # $P < .05$, 10^{-3} mol/L PBZ versus 10^{-5} mol/L PBZ, Pap/Lido, and control.

Incremental concentrations of NE also caused progressive vasoconstriction in control human RA segments (n = 11; $54.7\% \pm 7.5\%$ of maximal contraction to 30 mmol/L KCl; Figure 2). Soaking the RA in a combination of papaverine/lidocaine blood solution modestly but significantly attenuated this vasoconstriction response to 15 $\mu\text{mol/L}$ NE (n = 36; $35.6\% \pm 5.1\%$; $P = .04$). Although PBZ at 10 $\mu\text{mol/L}$ (n = 26) inhibited constriction to concentrations of NE greater than 7 $\mu\text{mol/L}$, PBZ at 1000 $\mu\text{mol/L}$ (n = 27) completely inhibited constrictor responses across all concentrations of NE (Figure 2). In summary, 1000 $\mu\text{mol/L}$ PBZ added to papaverine/lidocaine completely inhibits the vasoconstriction induced by PE and NE.

The potential for fasciotomy at the time of RA harvest to facilitate exposure of the vessel to PBZ pretreatment was investigated. At the highest concentration of PE tested (15 $\mu\text{mol/L}$), there was no significant difference between RA segments with fasciotomy and without fasciotomy with either papaverine/lidocaine treatment (n = 19 with fasciotomy; n = 13 without fasciotomy) or PBZ treatment (1000 $\mu\text{mol/L}$; n = 12 with fasciotomy; n = 10 without fasciotomy; Figure 3). Similarly, there was no benefit to fasciotomy with either papaverine/lidocaine pretreatment (n = 22 with fasciotomy; n = 14 without fasciotomy) or PBZ pretreatment (n = 17 with fasciotomy; n = 10 without fasciotomy) when vasoconstriction was achieved by NE (Figure 4).

RA endothelial function was tested by quantifying the relaxation response to increasing concentrations of ACh, a

Figure 2. Radial artery vasocontraction responses to increasing concentrations of norepinephrine, with or without pretreatment with 3 different concentrations of phenoxybenzamine (PBZ). PBZ attenuates vasocontraction to norepinephrine in a concentration-dependent manner. PBZ, Phenoxybenzamine pretreatment; Pap/Lido, papaverine/lidocaine pretreatment; control, no pretreatment. * $P < .05$, 10^{-3} mol/L PBZ versus 10^{-4} and 10^{-5} mol/L PBZ, Pap/Lido, and control. # $P < .05$, 10^{-3} mol/L PBZ versus 10^{-5} mol/L PBZ, Pap/Lido, and control.

Figure 3. Radial artery vasocontraction responses to 15 $\mu\text{mol/L}$ phenylephrine, with and without fasciotomy after treatment with 10^{-3} mol/L phenoxybenzamine or papaverine/lidocaine. PBZ, Phenoxybenzamine pretreatment; Pap/Lido, papaverine/lidocaine pretreatment. There were no significant differences between fasciotomy and no fasciotomy in the efficacy of PBZ or papaverine/lidocaine against 15 $\mu\text{mol/L}$ phenylephrine.

receptor-dependent stimulator of nitric oxide synthase. All RA segments had similar precontraction tensions generated by treatment with U46619 (PBZ-treated with fasciotomy, 14.4 ± 1.9 g of tension; PBZ-treated without fasciotomy, 15.0 ± 1.8 g; papaverine/lidocaine with fasciotomy, 13.1 ± 1.4 g; papaverine/lidocaine without fasciotomy, $14.9 \pm$

Figure 4. Radial artery vasocontraction responses to 15 $\mu\text{mol/L}$ norepinephrine, with and without fasciotomy after treatment with 10^{-3} mol/L phenoxybenzamine or papaverine/lidocaine. PBZ, Phenoxybenzamine pretreatment; Pap/Lido, papaverine/lidocaine pretreatment. There were no significant differences between fasciotomy and no fasciotomy in the efficacy of PBZ or papaverine/lidocaine against 15 $\mu\text{mol/L}$ norepinephrine.

1.6 g; $P = .79$). Endothelial function was not significantly attenuated in RA segments in which a fasciotomy was performed (Figure 5). In the segments treated with papaverine/lidocaine, those without fasciotomy ($n = 10$) demonstrated an $84.6\% \pm 6.8\%$ relaxation response to 12 $\mu\text{mol/L}$ ACh, and those with fasciotomy ($n = 17$) demonstrated an $80.7\% \pm 5.7\%$ relaxation response ($P = \text{not significant}$). In the segments treated with 1000 $\mu\text{mol/L}$ PBZ in addition to intraoperative papaverine/lidocaine, those without fasciotomy ($n = 9$) demonstrated an $81.0\% \pm 11.8\%$ relaxation response to 12 $\mu\text{mol/L}$ ACh, suggesting no additional impairment of endothelial function compared with RA segments treated with papaverine/lidocaine alone. Those that had fasciotomy and were treated with PBZ/papaverine/lidocaine ($n = 8$) showed a trend toward reduced endothelial function, averaging a $67.6\% \pm 5.2\%$ relaxation response to ACh ($P = .33$ compared with segments treated with PBZ/papaverine/lidocaine without fasciotomy; unpaired Student t test).

Discussion

This study showed that PBZ significantly attenuates the human RA constriction response to the clinically used vasopressors NE and PE in a dose-dependent manner. At a concentration of 1000 $\mu\text{mol/L}$, PBZ completely blocks vasocontraction to concentrations of α -adrenergic agents that are typically used during off-pump surgery to treat intraoperative hypotension and that are used after surgery to sustain systemic blood pressure. PBZ at concentrations of 100 and 10 $\mu\text{mol/L}$ has some efficacy in attenuating RA vasocontraction; however, these concentrations do not completely

block the response to NE and PE compared with 1000 $\mu\text{mol/L}$ PBZ, which completely blocked vasoconstriction but also unmasked the vasodilatation component of the mixed α - and β -adrenergic agent NE. Furthermore, performing an incision in the fascial plane to expose the graft to topical vasodilators does not increase the efficacy of either PBZ or papaverine/lidocaine. A pedicled harvest of the RA to avoid graft trauma, combined with pretreatment of the RA with 1000 $\mu\text{mol/L}$ PBZ to block vasospasm from α -adrenergic stimuli, may be important in preventing RA graft vasospasm and its complications, especially in the early postoperative period.

The optimal concentration at which PBZ completely abolished adrenergically induced vasoconstriction was 1000-fold greater than that reported for canine RA by Velez and colleagues.¹⁴ There is likely a species difference in response to α -adrenergic agents between human beings and dogs that may be related to the density of adrenergic receptors. In addition, the RAs arteries were skeletonized in the study by Velez and colleagues, and this may facilitate exposure to PBZ. It was thought that the presence of the soft tissue pedicle surrounding the human RA graft impedes contact of PBZ with the graft. However, in this study, increasing the exposure of the graft via fasciotomy did not change the efficacy of PBZ or papaverine/lidocaine. Complete skeletonization of the RA graft was not performed, as advocated by Taggart and colleagues,¹⁶ because skeletonization is contrary to the no-touch technique to avoid conduit trauma and may encourage vasospasm.

The optimal concentration used in this study is 6-fold less than that used by Taggart and colleagues.¹⁶ Concentrations of PBZ higher than 1000 $\mu\text{mol/L}$ are unnecessary because 1000 $\mu\text{mol/L}$ PBZ effectively blocks vasoconstriction by α -adrenergic agents. The formulation of PBZ used by both this study and that of Taggart and colleagues is a highly acidic solution composed of ethanol, hydrochloric acid, and propylene glycol that must be properly buffered to avoid endothelial or other cellular injury. The pH of Taggart and associates' blood/PBZ solution was not reported. However, Taggart and colleagues¹³ and Dipp and colleagues¹⁷ demonstrated full endothelial function with a blood-based solution of greater than 1000 $\mu\text{mol/L}$ PBZ. In this study, PBZ at a concentration of 1000 $\mu\text{mol/L}$ in a buffered crystalloid solution did not cause endothelial dysfunction. However, papaverine alone has been shown to cause a decrease in endothelium-dependent relaxation by ACh in internal thoracic arteries and RAs.¹⁷⁻²⁰ We observed that the use of papaverine/lidocaine in RA segments attenuated the endothelium-dependent relaxation response to ACh by 15% to 20% (data not shown). The use of papaverine in RA grafting is deleterious to the endothelium of the conduit; however, a properly buffered blood- or crystalloid-based

Figure 5. Radial artery endothelial function expressed as percentage relaxation to 12 $\mu\text{mol/L}$ acetylcholine, with and without fasciotomy after treatment with 10^{-3} mol/L phenoxybenzamine or papaverine/lidocaine. PBZ, Phenoxybenzamine pretreatment; Pap/Lido, papaverine/lidocaine pretreatment. There were no significant differences between treatments or fasciotomy.

solution of 1000 $\mu\text{mol/L}$ PBZ does not cause endothelial dysfunction.

External exposure alone via simple immersion of the graft in a PBZ solution may not be optimally effective. Because the vasa vasorum of the RA does not penetrate the muscular vessel media,²¹⁻²³ we also flushed the RA conduit intraluminally to maximize exposure of the graft to the agents.

In summary, pretreatment of the RA conduit with PBZ attenuates the vasoconstrictor response to the widely used vasopressors NE and PE. The efficacy of 1000 $\mu\text{mol/L}$ PBZ is not enhanced by increasing external exposure of the graft via an incision in the soft tissue pedicle (fasciotomy); however, gentle intraluminal flushing is important for delivering the drug to the muscular vessel media. RA vasospasm is effectively blocked ex vivo by carefully preserving the soft tissue pedicle protecting the graft and pretreating the conduit with PBZ. Endothelial function is preserved with the use of PBZ. Therefore, we recommend the no-touch technique of RA harvesting and brief pretreatment of the RA graft with 1000 $\mu\text{mol/L}$ PBZ.

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References

1. Carpentier A, Guermontprez JL, Deloche A, Frechette C, DuBost C. The aorta-to-coronary radial artery bypass graft: a technique avoiding pathologic changes in grafts. *Ann Thorac Surg*. 1973;16:111-21.
2. Carpentier A. Discussion of Geha AS, Krone RJ, McCormick JR, Baue

- AE. Selection of coronary bypass: anatomic, physiological, and angiographic considerations of vein and mammary artery grafts. *J Thorac Cardiovasc Surg.* 1975;70:414-31.
3. Acar C, Jebara VA, Portoghese M, Beyssen B, Pagny JY, Grare P, et al. Revival of the radial artery for coronary artery bypass grafting. *Ann Thorac Surg.* 1992;54:652-60.
 4. Acar C, Ramshevi A, Pagny J-Y, Jebara V, Barrier P, Fabiani J-N, et al. The radial artery for coronary artery bypass grafting: clinical and angiographic results at five years. *J Thorac Cardiovasc Surg.* 1998; 116:981-9.
 5. Califore AM, Di Giammarco G, Luciani N, Maddestra N, Di Nardo E, Angelini R. Composite arterial conduits for a wider arterial myocardial revascularization. *Ann Thorac Surg.* 1994;58:185-90.
 6. Dietl C, Benoit C. Radial artery grafts for coronary artery bypass grafting. *Ann Thorac Surg.* 1995;60:102-10.
 7. Brodman RF, Frame R, Camacho M, Hu E, Chen A, Hollinger I. Routine use of unilateral and bilateral radial arteries for coronary artery bypass surgery. *J Am Coll Cardiol.* 1996;28:959-63.
 8. da Costa FDA, da Costa IA, Poffo R, Abuchaim D, Gaspar R, Garcia L, et al. Myocardial revascularization with the radial artery: a clinical and angiographic study. *Ann Thorac Surg.* 1996;62:475-80.
 9. Manasse E, Sperti G, Suma H, Canosa C, Kol A, Martinelli L, et al. Use of the radial artery for myocardial revascularization. *Ann Thorac Surg.* 1996;62:1076-83.
 10. Bahn A, Gupta V, Choudhary SK, Sharma R, Singh B, Aggarwal R, et al. Radial artery in CABG: could the early results be comparable to internal mammary artery graft? *Ann Thorac Surg.* 1999;67:1631-6.
 11. Apostolidou IA, Skubas NJ, Despotis GJ, Kallinteri E, Hogue CW, Lappas DG, et al. Occurrence of myocardial ischemia immediately after coronary revascularization using radial artery conduits. *J Cardiothorac Vasc Anesth.* 2001;15:433-8.
 12. Gabe ED, Figal JC, Wisner JN, Laguens R. Radial artery graft vasospasm. *Eur J Cardiothorac Surg.* 2001;19:102-4.
 13. Taggart DP, Dipp M, Mussa S, Nye PCG. Phenoxybenzamine prevents spasm in radial artery conduits for coronary artery bypass grafting. *J Thorac Cardiovasc Surg.* 2000;120:815-7.
 14. Velez DA, Morris CD, Muraki S, Budde JM, Otto RN, Zhao Z-Q, et al. Brief pretreatment of radial artery conduits with phenoxybenzamine prevents vasoconstriction long term. *Ann Thorac Surg.* 2001;72:1977-84.
 15. Johnson WH III, Cromartie RS III, Arrants JE, Wuamett JD, Holt JB. Simplified method for candidate selection for radial artery harvesting. *Ann Thorac Surg.* 1998;65:1167.
 16. Taggart DP, Mathur MN, Ahmad I. Skeletonization of the radial artery: advantages over the pedicled technique. *Ann Thorac Surg.* 2001;72:298-9.
 17. Dipp MA, Nye PCG, Taggart DP. Phenoxybenzamine is more effective and less harmful than papaverine in prevention of radial artery vasospasm. *Eur J Cardiothorac Surg.* 2001;19:482-6.
 18. Cooper GJ, Wilkinson GAL, Angelini GD. Overcoming perioperative spasm of the internal mammary artery: which is the best vasodilator? *J Thorac Cardiovasc Surg.* 1992;104:465-8.
 19. Cooper GJ, Gillot T, Parry EA, Kennedy A, Wilkinson GA. Papaverine injures the endothelium of the internal mammary artery. *Cardiovasc Surg.* 1995;3:553-5.
 20. He G-W. Verapamil plus nitroglycerin solution maximally preserves endothelial function of the radial artery: comparison with papaverine solution. *J Thorac Cardiovasc Surg.* 1998;115:1321-7.
 21. He G-W. Arterial grafts for coronary artery bypass grafting: biological characteristics, functional classification, and clinical choice. *Ann Thorac Surg.* 1999;67:277-84.
 22. van Son JAM, Smedts F, Vincent JG, van Lier HJJ, Kubat K. Comparative anatomic studies of various arterial conduits for myocardial revascularization. *J Thorac Cardiovasc Surg.* 1990;99:703-7.
 23. Chester AH, Amrani M, Borland JAA. Vascular biology of the radial artery. *Curr Opin Cardiol.* 1998;13:447-52.